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4 **Identification of Mosaic *Neisseria gonorrhoeae* Penicillin-Binding Protein 2 — 2008, San**

5 **Francisco, California**

6 Running title: *N. gonorrhoeae* mosaic PBP2 in San Francisco

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1 **Abstract:**

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3 Using a real-time PCR assay specific for a mosaic *penA* allele that has been associated with oral
4 cephalosporin resistance in Asia, fifty-four available *Neisseria gonorrhoeae* isolates collected in
5 San Francisco from January–October, 2008 were analyzed. Five isolates tested positive for the
6 mosaic *penA* gene by real-time PCR. DNA Sequencing revealed two mosaic *penA* alleles (SF-A
7 and SF-B). Isolates with SF-A and SF-B alleles possessed elevated MIC for the oral
8 cephalosporins cefpodoxime and cefixime.

9

1 Isolates with decreased susceptibility to third-generation cephalosporins, particularly oral
2 cephalosporins, have emerged in Asia, Australia, and elsewhere (1, 2, 4, 5, 7, 11, 12). Initial
3 reports linked this decreased susceptibility to oral cephalosporins to an altered, mosaic penicillin
4 binding protein 2 (PBP2) coded by the *penA* gene characterized by multiple genetic changes with
5 segments that are nearly identical to the homologous regions of the *penA* genes of related
6 commensal *Neisseria* species (2,3). Recently a real-time PCR assay has been developed for
7 detection of this mosaic *penA* gene (8). We used this real-time PCR assay to determine whether
8 the mosaic *penA* allele is present in clinical isolates of *Neisseria gonorrhoeae* in San Francisco.
9
10 Fifty-four *Neisseria gonorrhoeae* isolates collected during January–October 2008 from male
11 patients with symptomatic urethritis were available for testing. Of these, 5 isolates were found to
12 be reactive by real-time PCR for the mosaic *penA* gene (SM-1, SM-2, SM-3, SM-4 and SM-5).
13 An assortment of 100 *N. gonorrhoeae* isolates from 2002–2006 collected in San Francisco were
14 also analyzed with the same real-time PCR assay, and none of those specimens were found to be
15 reactive.
16
17 In order to confirm the presence of a mosaic *penA* allele in the five real-time PCR-reactive
18 isolates, the *penA* genes of these isolates were analyzed by DNA sequencing. The primers used
19 for the amplification and sequencing of the *penA* genes are shown in Table 2. As shown in
20 Figure 1, two distinct *penA* alleles were found in the five PCR-positive isolates. These two
21 alleles were designated “SF-A” (3 isolates: SM-1, SM-2, and SM-3) and “SF-B” (2 isolates:
22 SM-4 and SM-5). We compared these two novel *penA* alleles (SF-A and SF-B) to both a wild-
23 type *penA* allele (GenBank # M32091) and the mosaic *penA* allele associated with oral

1 cephalosporin resistance in Asia and Australia, AB071984 (2). Neither of the San Francisco
2 mosaic alleles was found to contain all of the mutations associated with AB071984. The
3 translated amino acid sequence of SF-A is identical to that of AB071984 for the first 549 amino
4 acid residues. From amino acid 550 to the end of the translated sequence, the SF-A allele is
5 identical to the reference wild type allele. The translated amino acid sequence of SF-B possesses
6 greater dissimilarity to AB071984 than SF-A. Although containing many of the mosaic-
7 associated mutations, SF-B lacks AB071984-associated codons at amino acids 279, 285, 288 and
8 291. Additionally, SF-B lacks all of the AB071984-specific codons from codon 388 to the end of
9 the translated amino acid sequence. Interestingly, SF-B possessed unique amino acid residues
10 distinct from wild type, SF-A and AB071984 at residues 35, 42, 70, 230 and 515.

11
12 The susceptibilities of the five real-time PCR positive isolates to certain third-generation
13 cephalosporins were evaluated. Results of agar dilution susceptibility testing were available
14 through the Gonococcal Isolate Surveillance Program (GISP) for ceftriaxone. Isolates SM-1 and
15 SM-2 each possessed a ceftriaxone MIC of 0.06 ug/mL, and isolate SM-3 had a ceftriaxone MIC
16 of 0.03 μ g/mL (Table 1). SM-4 and SM-5 each possessed ceftriaxone MICs of \leq 0.008 μ g/mL.
17 All five isolates were evaluated with regard to their susceptibilities to the oral third-generation
18 cephalosporins, cefixime and cefpodoxime, using agar dilution (protocol available at
19 http://www.cdc.gov/std/gisp/protocol2006_web_version_rev12_2007.pdf Accessed 05/01/2008).
20 The five *penA* mosaic isolates were compared with two isolates from San Francisco determined
21 by real-time PCR and *penA* sequencing to possess non-mosaic *penA* alleles (SW-1 and SW-2).
22 Isolates with the SF-A *penA* allele (SM-1, SM-2 and SM-3) had MIC values for both cefixime
23 and cefpodoxime that were notably higher than the values for strains found containing wild-type

1 *penA* alleles. SF-B-containing isolates possessed modestly elevated MIC values to cefpodoxime,
2 while possessing little or no elevation in MIC to cefixime compared with strains with non-
3 mosaic *penA* alleles. Further investigation of the five isolates with mosaic *penA* alleles included
4 multi-antigen sequence typing of these five strains (NG-MAST) using a previously published
5 method (6) Four of the five isolates possessed the NG-MAST sequence type 1407 (Table 1). SM-
6 2 possessed sequence type 1513.

7 These data demonstrate the presence of two previously undescribed *penA* alleles (SF-A and SF-
8 B) within *Neisseria gonorrhoea* associated with elevated cephalosporin MICs in San Francisco.
9 These alleles resemble the mosaic *penA* alleles previously associated with cephalosporin
10 resistance in Asia and Australia (2, 5, 11). Although the exact relationship between the mosaic
11 *penA* and the development of decreased susceptibility to cephalosporins is not completely
12 understood, these findings are concerning because they might indicate impending development
13 and spread of isolates in the United States resistant to cephalosporins, particularly oral third-
14 generation cephalosporins.

15

16 Of the two newly described alleles, SF-A most resembles the previously described mosaic allele
17 associated with strains resistant to oral third-generation cephalosporin. SF-A also has 99%
18 similarity with *penA* allele from an *N. gonorrhoeae* isolate with cefuroxime (a second generation
19 cephalosporin) resistance (Gen Bank # DQ335216, unpublished submission, JE Corkill). Osaka
20 et al identified three amino acid alterations important for oral cephalosporin resistance, I312M,
21 V316T, and G545S among cephalosporin resistant isolates in Japan (9). SF-A and SF-B both
22 have I312M and V316T, but only SF-A has G545S. Both isolates bearing the SF-B *penA* allele
23 were found to have lower cephalosporin MICs than isolates with the SF-A allele. Interestingly,

1 isolates with the either the SF-A or the SF-B alleles had elevated cefpodoxime MICs, while SF-
2 A had a higher MIC for cefixime than SF-B. Both the *penA* allele type and the presence of
3 I312M, V316T, and G545S appeared to correlate with the degree to which cephalosporin MICs
4 were elevated. These data support previous reports demonstrating the importance of these codons
5 in cephalosporin resistance.(9)
6
7 These results raise several questions for future study including determining whether isolates with
8 SF-A or SF-B *penA* alleles are associated with treatment failure. The primary method of *N.*
9 *gonorrhoeae* detection in our laboratory includes nucleic acid amplification testing which does
10 not involve the collection of viable organisms for isolation. We are currently working to develop
11 an assay that will allow us to screen such specimens for the presence of mosaic *penA* alleles in an
12 effort to more carefully define the prevalence of these alleles in our setting and identify patients
13 for close follow-up after treatment.

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5 **Figure Legends:**

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7 Figure 1: Comparison of *penA* gene sequences for specimens with mosaic *penA* alleles detected

8 in this study (SF-A and SF-B) to wild type sequence (GenBank #M32091) and a previously

9 described mosaic *penA* allele (GenBank #AB071984).

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Table 2. Oligonucleotides utilized for amplification and sequencing of *penA* genePCR Primers:

Fragment 1 of *penA* gene:

5'-GCATCAGGATAATAATAACGAGAAG-3' *

5'-TGTAAGGCAGGGTATTGAAT-3'

Fragment 2 of *penA* gene:

5'-TCGGGCAATACCTTTATGGTGG AACAT-3' **

5'-CAGCCAAAGGGGTAACTTGCTGAAC-3' **

Sequencing primers:

5'-GCATCAGGATAATAATAACGAGAAG-3' *

5'-TGTAAGGCAGGGTATTGAAT-3'

5'-AACCTTCCTGACCTTTGCCGTC-3' †

5'-AAAACGCCATTACCCGAAGGG-3' †

5'-CAGCCAAAGGGGTAACTTGCTGAAC-3' **

5'-AATTGAGCCTGCTGCAATTGGC-3' †

5'-GTTGGATGCCCGTACTGGG-3'. ††

* Reference 10

** Reference 2

† Reference 3

†† Reference 8

Table 1. Description of *Neisseria gonorrhoeae* isolates with mosaic *penA* alleles in San Francisco, CA, 2008

Isolate	Agar Dilution			Mosaic PCR ^a	<i>penA</i> allele	NG-MAST ST ^b
	Cro ($\mu\text{g/mL}$)	Cfm ($\mu\text{g/mL}$)	Cpd ($\mu\text{g/mL}$)			
SM-1	0.060	0.25	1.0	(+)	SF-A	1407
SM-2	0.060	0.125	0.5	(+)	SF-A	1513
SM-3	0.030	0.25	1.0	(+)	SF-A	1407
SM-4	≤ 0.008	≤ 0.008	0.06	(+)	SF-B	1407
SM-5	≤ 0.008	0.015	0.25	(+)	SF-B	1407
SW-1	≤ 0.008	≤ 0.008	0.015	(-)	NM	242*
SW-2	≤ 0.008	0.015	0.03	(-)	NM	2992

NM: Non-mosaic *penA* sequence, Cro: ceftriaxone, Cfm: cefixime, Cpd: cefpodoxime,

NG-MAST: *Neisseria gonorrhoeae* multiantigen sequence typing, ST: sequence type

^a reactivity with real-time PCR (8)

^b sequence type (determined by NG-MAST (6))

* isolate not previously identified by NG-MAST database; 99% similarity to ST242

Figure 1.

	10	20	30	40	50	60	70	80	90	100
M32091	PBP2MLIKSEYKPRMLPKKEQVKKPMTSNGRISFVLMAMAVLFACLIARGLYLQTVTYNFKKEQGDNRIVRTOALPATRGTVSDRNGAVLALSAPTESLFAVPK									
SF-A									
SF-BI.....G.....T.....									
AB071984									
	110	120	130	140	150	160	170	180	190	200
M32091	PBP2DMKEMPSAAQLERLSELVDVPVDVLRNKLEQKGSFIWIKRQLDPKVAEEVKALGLENFVFEKELKRHYPMGNLFAHVIGFTDIDGKGQEGLELSLEDLSL									
SF-A	E.....A.....S.....									
SF-B	E.....									
AB071984	E.....A.....S.....									
	210	220	230	240	250	260	270	280	290	300
M32091	PBP2YGEDGAEVVLRDRQGNIVDSLDSPRNKAPQNGKDIILSLDQRIQTLAYEELNKAVEYHQAKAGTVVLDARTGEILALANTPAYDPNRPGRADSEQRNR									
SF-A	HAGE.....E.....V.....E..K..Q.....									
SF-B	HAGE.....K.....									
AB071984	HAGE.....E.....V.....E..K..Q.....									
	310	320	330	340	350	360	370	380	390	400
M32091	PBP2AVTDMIEPGSAIKPFVIAKALDAGKTDLNERLNTQPYKIGPSPVRDTHVYPSLDVRGIMQKSSNVGTSKLSARFGAEEMYDFYHELIGIVRMHSGFPGET									
SF-AM..T.....S..V.ATDTF..L.....SAT.Q.....T.....M.TPK.....D..V.....									
SF-BM..T.....S..V.ATDTF..L.....AT.Q.....T.....M.TPK.....D..V.....									
AB071984M..T.....S..V.ATDTF..L.....SAT.Q.....T.....M.TPK.....D..V.....									
	410	420	430	440	450	460	470	480	490	500
M32091	PBP2AGLLRNWRRWRPIEQATMSFGYGLQLSLQLARAYTALTHDGVLLPLSFEKQAVAPQGKRIFKESTAREVNRNLMVSVTEPGGTGTAGAVDGFVDAKTGT									
SF-A	...S...QK.....V...E..V.....K..VI.A..KK..E.....A.....									
SF-BV.....									
AB071984	...S...QK.....V...E..V.....K..VI.A..KK..E.....A.....									
	510	520	530	540	550	560	570	580		
M32091	PBP2ARKFVNGRYADNKHVATFIGFAPAKNPRVIVAVTIDEPTAHGGYGGVVAGPPFKKIMGGSNLNIGISPTKPLT-AAAVKTPS									
SF-A	...L...V.Y.....N...S.....									
SF-BI.....									
AB071984	...L...V.Y.....N...S...T..V..QV.....V.....NV.....									